

## ***Pseudomonas* CULTURE TECHNIQUE**

# Production and Application of Syringomycin E as an Organic Fungicide Seed Protectant against *Pythium* Damping-off

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### Abstract

Syringomycin E (SRE) is a cyclic lipodepsinonapeptide with potent antifungal activity and is produced by certain strains of *Pseudomonas syringae* *pv.* *syringae*. In this study, its potential as an organic-compatible agrofungicide and vegetable seed treatment against the soilborne pathogen *Pythium ultimum* *var. ultimum* was examined. A variant of *P. syringae* *pv. syringae* strain B301D with enhanced SRE-producing capabilities was isolated and grown in a bioreactor with SRE yields averaging 50 mg/l in 40 h. SRE was extracted and purified through a large-scale chromatography system using organic-compatible processes and reagents. Organiccompatible and scalably produced SRE is potentially a novel organic fungicide seed protectant.



**Winpact Model: FS-02-B10P**

### Introduction

Strains of *Pseudomonas syringae* *pv. syringae* are epiphytes and opportunistic pathogens on healthy and susceptible plants, respectively. They produce two classes of non-ribosomal cyclic lipodepsipeptides commonly known as syringomycins and syringopeptins. Syringomycin E (SRE) is an abundant Syringomycin analog with a 3- hydroxydodecanoic acid group attached to a nonapeptide ring (Segre et al. 1989). Antifungal activities of SRE have been reported against many fungal species, including soilborne pathogens such as *Pythium*, *Fusarium* and *Rhizoctonia* (De Lucca et al. 1999).

As a secondary metabolite natural product, SRE is a potential natural fungicide for disease control applications. SRE inhibits oosporic phytopathogens such as *Pythium* sp. making it attractive to explore as a seed protectant against diseases caused by these pathogens.

*Pseudomonas syringae* *pv. syringae* B301D is principally used for the production of SRE in potato dextrose broth (PDB) medium (Zhang and Takemoto 1987), syringomycin minimal (SRM) broth (Gross 1985; Mo and Gross 1991) or improved minimal medium (IMM) broth (Surico et al. 1989), and the methods provide SRE levels in milligram quantities after several days of growth, performed typically without agitation.

The goals of this study were to (i) produce scalable amounts of SRE by methods that allow qualification for organic use certification.

## Materials and Methods

### Medium modification

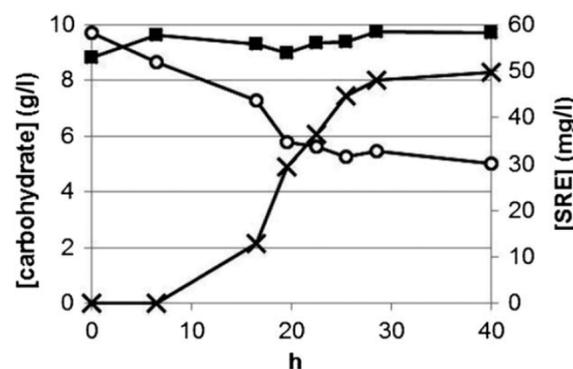
Modifications of growth medium, SRM (1% w/v glucose, 0.4% w/v histidine, 0.8 mM MgSO<sub>4</sub>, 1 mM FeCl<sub>3</sub> and 0.8 mM potassium phosphate, pH7) (Gross 1985; Mo and Gross 1991), were performed to improve SRE production by *P. syringae pv. syringae*. The antifungal activities of *P. syringae pv. syringae* strain 2B6 culture extracts were examined as a function of different carbon compounds substituted for 1% (w/v) glucose in SRM. Tested were glycerol, ribose, galactose, mannitol, lactic acid, acetate, or succinate, each at 1% (w/v), glucose plus mannitol, each at 1 or 2% (w/v), glucose, 1% (w/v), plus fructose, 0.1% (w/v), and arbutin, 100 μM, and glucose, 1% (w/v), plus fructose, 0.1% (w/v), and salicylate, 100 μM. The inclusion of 1% (w/v) glucose plus 1% (w/v) mannitol gave the highest antifungal activity, and this mannitol-supplemented SRM was named ‘improved SRE medium’ or ‘ISM’. SRE production by *P. syringae pv. syringae* strain G10 was examined with growth in SRM, ISM, SRM plus 100 μM arbutin and 0.1% (w/v) fructose, ISM plus 100 μM arbutin and 0.1% (w/v) fructose, and SRM plus 100 μM salicylate and 0.1% (w/v) fructose in agitated flask cultures.

### Scalable SRE production

*Pseudomonas syringae pv. syringae* strain G10 was initially grown in 500 ml of ISM with rotary shaking (200 rpm) at 28°C for 15 h. The 500 ml culture was used as inoculum for a 10 l ISM culture in a **10 L working volume Winpact Bioreactor and Fermenter System (Major Science, Saratoga, CA, USA)** with conditions maintained at pH 6.7, 28°C, 2.5 slpm aeration and 350 rpm stirring. At 40 h after inoculating the bioreactor, SRE was only detected (by HPLC) in the foam that accumulated above the culture medium and not in the culture broth. SRE was collected by siphoning the foam material.

## Results

In an agitated and aerated **10 L capacity Winpact bioreactor** and with ISM as growth medium, *P. syringae pv. syringae* strain G10 grew and produced foam on the culture surface. In a typical 40-h bioreactor SRE production run, glucose was consumed while mannitol was not (Fig. 1). The bioreactor culture began to foam on the liquid surface between 15 and 30 h coincident with initial detection of SRE in the culture broth (at 0.036–0.06 mg/l SRE). At 40 h, SRE was detected in the foam above the culture broth but not in the culture broth. The SRE yield averaged ~50 mg/l of culture.



**Fig. 1** *Pseudomonas syringae pv. syringae* strain G10 growing in ISM in a Winpact Bioreactor and Fermenter system with aeration and agitation produces SRE (x) and consumes glucose (O). Mannitol (■) is not consumed during growth.

## References

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